

Moving Insert to iGEM Vector

Restriction Digestion to Remove Insert from PGEM-T

<u>Reagent</u>	<u>Volume (ul)</u>
Miniprep product (500 ng/ul)	1*
10x BSA	2.5
10x Buffer 3	2.5
EcoRI (undiluted)	0.5**
PstI (undiluted)	0.5**
H2O	12.5

	25 ul

Incubate 37 deg Celsius; 1 - 2 hours

Incubate 65 C, 20 minutes

*The final concentration of the DNA in this reaction should be 20ng/ul. The volume of DNA (minprep product) you add to your reaction will be dependent upon the concentration of that DNA. For example: 250 ng/ul x 2ul = 500ng... 500ng/25ul = 20ng/ul). If you have more or less DNA, the volume added should be adjusted such that the final concentration of DNA in the digestion is 20 ng/ul. If you have questions about this – ask! *If your DNA is under 40ng/ul, concentrate it before digesting.*

**Can you accurately pipette 0.5 ul with the pipettes you're using? No; so make a cocktail. Only digesting one reaction? Instead of pipetting directly into the reaction, mix equal small volumes of EcoRI and PstI together, then use 1ul of that. Alternatively, make a cocktail for two reactions and only use half of it.

Example:

- Miniprep product concentration = 100ng/ul
Use 5ul product to obtain, and adjust the water, such that the total reaction volume is still 25 ul
 $100\text{ng/ul} \times 5\text{ul} = 500\text{ng} \dots 500\text{ng}/25\text{ul} = 20\text{ng/ul}$
- Or, work backwards:
Desired concentration: 20ng/ul in 25ul ; $(20\text{ng/ul}) \times (25\text{ul}) = 500\text{ng}$
Concentration of your DNA: 100ng/ul; $(500\text{ng}) / (100\text{ng/ul}) = 5\text{ul}$
Check your units – they should all cancel to leave you only with the units you're going after.
$$\frac{(\text{Desired concentration of DNA ng/ul}) \times (\text{Volume of reaction ul})}{(\text{Concentration of your DNA ng/ul})} = \text{Volume to use (ul)}$$

Ligation into predigested iGEM Vector

Equal masses of insert and vector should be added to the ligation mix. The iGEM vectors that are currently predigested are at 20ng/ul and your Miniprep digest is at 20ng/ul.

<u>Reagent</u>	<u>Volume (ul)</u>
2X Ligase Buffer	10
Miniprep digest (20 ng/ul)	5
Digested Vector (Kan Resistance) (20 ng/ul)	5
T4 DNA Ligase	1

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Incubate 4C, overnight

Transform 10 ul of ligation into 50ul DH5 α Competent Cells

Plate cells on LB Kan (not LB Carb!)

Incubate 37C overnight

Check colonies for insert of correct size (and the correct vector) by miniprep and digest.