Plasmid Mini-Preps using Qiagen solutions

Recommend doing in batches of 12 (or multiples thereof)

1. Inoculate your colony into 1.5 mL sterile LB in a sterile glass tube using a long stick to inoculate (no drug is OK, drug containing is also OK).

2. Grow up overnight at 37 degrees, shaking or rolling is not needed.

3. Transfer culture to microfuge tube and spin down 1.5 ml of overnight bacterial culture (1 min, top speed). Remove as much supernatant as possible.

4. Resuspend in 200 microliters of Qiagen's P1 buffer containing RNase A by vortexing.

3. **Immediately** add 200 microliters of room temperature P2 (lysis) solution and mix by inversion (**do not vortex**).

4. **Immediately** add 200 microliters of cold P3 (neutralization) buffer and mix by inversion (**do not vortex**).

5. **Immediately** spin in microcentrifuge, top speed, 10 minutes at 4^0 (room temperature for 5 min. works also).

6. Carefully pour supernatant (hopefully without dislodging yucky protein pellet) into a new tube containing 0.5 ml (approximately 1 volume) of isopropanol. Mix.

7. **Immediately** centrifuge DNA precipitate 5 minutes top speed at room temperature. Discard supernatant.

8. Re-centrifuge 10 seconds and remove remaining supernatant with micropipette.

9. Rinse DNA pellet with 100 microliters of 70% EtOH.

10. Centrifuge pellet 10 secondstop speed and remove supernatant with micropipette.

11. Air dry pellet 30 min or until completely dry and resuspend in 20 microliters of TE. Use 0.5 microliter of this DNA in a 10-20 microliter restriction enzyme digest.